# Efficacy of Standard Methods for Water Testing at a Range of Nonstandard Temperature

## Background

Standard, reliable testing for fecal contamination of water remains inaccessible to the majority of the world's population. Evaluation of water sources in developing countries (and following emergencies or natural disasters) cannot rely on electricity, skilled personnel, or a controlled laboratory environment. A simple, specific, and reliable field test that can be used in a variety of conditions is urgently needed.





### Materials and Methods

In an effort to test the viability of currently available *Escherichia coli* detection assays at an assortment of temperatures, 15 strains of *E. coli* from various geographical areas, animal, and soil sources were used. Each was inoculated onto 5 standard and nonstandard tests and incubated at 5 temperatures: 23, 30, 37, 40, and 45°C. Tests studied included Petrifilm™, Colilert®, EPA method 1604 (MI Medium), MacConkey II (MAC II) Agar and EC Medium multiple-well fermentation.

#### **Assays Tested**

- Two standard ß-glucuronidase assays—Colilert\* (IDEXX, Westbrook, ME) and membrane filtration with MI Agar (BD-BBL, Sparks, MD).
- Two nonstandard (and less expensive) assays using 4-methylumbelliferyl-p-glucuronide (MUG) as the indicator nutrient. EC Medium with MUG (Difco, Sparks, MD) was chosen for comparison with Colilert\*, and Mac II Agar with MUG (BD, Sparks, MD) was used in contrast with MI Agar.
- A Petrifilm<sup>™</sup> (3M<sup>™</sup> Microbiology, St. Paul, MN) assay was also included.

#### Protocol

All test protocols were adapted from the standard methods recommended by either the manufacturer or the EPA. The Colilert® assay consisted of three snap packs (pre-measured for 100 mL samples) combined within 300-mL of E. coli-inoculated water, and divided by 1 mL samples into five 48-well culture plates. Similarly, 11.1 g of dry EC Medium were added to a 300-mL sample and transferred to 48-well plates. Prior to experimentation, the EC Medium was tested repeatedly for sterility by incubation of uninoculated medium at 37°C for 24 hours. Petrifilm™ consisted of 1-mL water samples transferred onto five Coliform Count Plates. For the agar assays, a total of ten 100-mL samples were run through a 0.45-µm filter and transferred to agar medium. MI Agar and MAC II Agar were purchased preplated, and alcohol-sterilized forceps were used in handling of the filters.

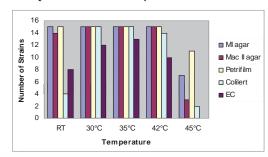
#### E. Coli Strains Tested

| Accession# | Strain Name | 0   | н  | K | Class               | Host               | Locale      | Date | Clinical |
|------------|-------------|-----|----|---|---------------------|--------------------|-------------|------|----------|
| TW02282    | MT-512      | 2   |    |   |                     | chicken            | France      | 1972 | trachea  |
| TW02283    | MT-513      | 2 2 |    |   |                     | chicken            | France      | 1972 | salphins |
| TW02284    | MT-515      | 78  |    |   |                     | chicken            | France      | 1972 | lung     |
| TW06930    | 92-1232     | 141 | 12 |   |                     | goose              | USA (NY)    |      | feces    |
| TW03284    | ECOR-21     | 121 |    |   |                     | steer              | Bali        |      | healthy  |
| TW03289    | ECOR-29     | 150 | 21 |   |                     | k-rat              | USA (NV)    |      | healthy  |
| TW02053    | ECOR-33     | 7   | 21 |   | ECORr5              | sheep              | USA (CA)    |      | healthy  |
| TW03292    | ECOR-34     | 88  | NM |   | ECOR <sub>1</sub> 5 | dog                | USA (MA)    |      | healthy  |
| TW02060    | ECOR-45     | ON  | M  |   |                     | pig                | Indonesia   |      | healthy  |
| TW03298    | ECOR-47     | OM  | 18 |   |                     | sheep              | New Guinea  |      | healthy  |
| TW02066    | ECOR-67     | 4   | 43 |   |                     | goat               | Indonesia   |      | healthy  |
| TW11566    | MT2a        |     |    |   |                     | environment (soil) | Puerto Rico |      |          |
| TW11569    | MT4a        |     |    |   |                     | environment (soil) | Puerto Rico |      |          |
| TW11573    | MT7c        |     |    |   |                     | environment (soil) | Puerto Rico |      |          |

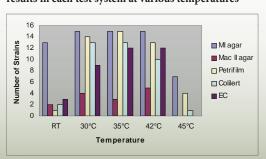
#### Results

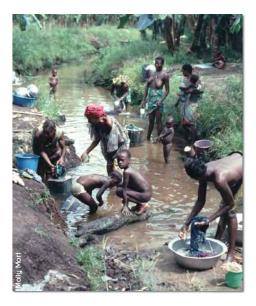
- *E. coli* can be cultured at temperatures ranging from approximately 22°C to 40°C. Some strains could be cultured at 45°C, but not all. Preliminary testing at 4°C demonstrated no visible growth at 24 hours.
- The commercial tests and media used have been optimized for use at 35°C ± 2°C. However, growth was observed at temperatures from 22°C to 42°C using these tests.
- Of the tests studied, membrane filtration with MI Agar gave the best performance for β-glucuronidase detection, β-galactosidase detection, and *E. coli* specificity across all temperatures. Petrifilm™ produced the best growth of *E. coli* at all temperatures but lacked specificity. Colilert® did not perform well outside its optimized temperature range so would be of limited usefulness in low-resource settings. The nonstandard tests, Mac II Agar and EC Medium multiple-well fermentation, gave inconsistent results and would not be useful for a reliable test in uncontrolled conditions.

## Number of strains exhibiting growth at 5 temperatures in each test system



# Number of strains that exhibited *E. coli*-positive results in each test system at various temperatures





#### Conclusions

- *E. coli* can be cultured at temperatures from 22°C to 40°C. However, 45°C gave inconsistent results strain to strain.
- This range would allow testing using some standard methods without use of an incubator to test for fecal contamination of drinking water supplies.
- MI agar gave the best specificity for E. coli, but Petrifilm gave comparable results and is more easily transportable to the field, easy to use and cost-effective.

#### Acknowledgements

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www.path.org

#### References

Carolina Biological Supply Company, 3M" Petrifilm" Coliform Count Plates Instruction Manus Burlington, NC: Carolina Biological Supply Company.

EPA. Approved Methods for Microorganisms. 2006. Ground Water and Drinking Water. Available

EPA. Method 1604: Total Coliforms and Escherichia coli in Water by Membrane Filtration Using

dexx Laboratories. Colilert\* Test Kit. Westbrook, ME: Idexx Laboratories; 200

Clasen T, Smith L. The Drinking Water Response to the Indian Ocean Tsunami Including the Role of

Jnited Nations System. Pakistan 2005 Earthquake: Early Recovery Framework. Islamabad, Pakist: Jnited Nations System; 2005.

UNESCO. 2003. Facts and Figures. International Year of Freshwater. www.wateryear2003.org

WHO/UNICEF. Meeting the MDG Drinking Water and Sanitation Target: A Mid-Term Assessme of Progress. Geneva, Switzerland: WHO; 2004.

